

## The Long-lasting Antiproliferative Effect of 15-Deoxyspergualin through its Spermidine Moiety

MANABU KAWADA, TETSUYA SOMENO, HIRONOBU IINUMA,  
TOHRU MASUDA, MASAOKI ISHIZUKA\* and TOMIO TAKEUCHI

Institute for Chemotherapy, M. C. R. F.,  
18-24 Miyamoto, Numazu-shi, Shizuoka 410-0301, Japan

(Received for publication February 16, 2000)

15-Deoxyspergualin (DSG) inhibited growth of mouse EL-4 lymphoma cells with an  $IC_{50}$  0.02  $\mu\text{g/ml}$ . Even though the cells were treated with DSG for only 4 hours and then washed, the antiproliferative effect lasted long with an  $IC_{50}$  0.4  $\mu\text{g/ml}$ . DSG has spermidine and guanidine moieties in its structure. One decomposed element containing guanidine moiety inhibited the growth at higher doses than DSG, but the effect did not last long unlike DSG. While another element containing spermidine moiety did not affect the growth, it diminished the long-lasting antiproliferative effect of DSG by pretreatment of the cells. Pretreatment with polyamines such as putrescine, spermidine, and spermine also diminished the effect of DSG. Furthermore, *N*-alkylation of spermidine moiety in DSG abolished the antiproliferative effect. These results suggested that DSG binds to the cells through its spermidine moiety and exerts its long-lasting antiproliferative effect.

15-Deoxyspergualin (DSG) is the most potent synthetic analogue of spergualin, which was isolated as an antitumor compound from microbial cultured broth (Fig. 1).<sup>1-3</sup> DSG exhibits antitumor effect on various tumor cell lines *in vitro* and *in vivo*.<sup>4,5</sup> Furthermore, DSG has a potent immunosuppressive effect and has been used as an immunosuppressant.<sup>6,7</sup> Many efforts have been made on elucidating the mechanism for DSG. It is reported that DSG binds to Hsc70 and inhibits the function of Hsc70.<sup>8</sup> In respect to the antitumor effect, DSG is reported to inhibit the cell cycle progression at G1 phase<sup>9,10</sup> and tumorigenic angiogenesis.<sup>11,12</sup> However, the precise mechanism for DSG action is still unclear. DSG is composed of spermidine and guanidine moieties. In this paper we focused on the structural characteristics of DSG and studied the mechanism for DSG action on tumor cell growth.

### Materials and Methods

#### General

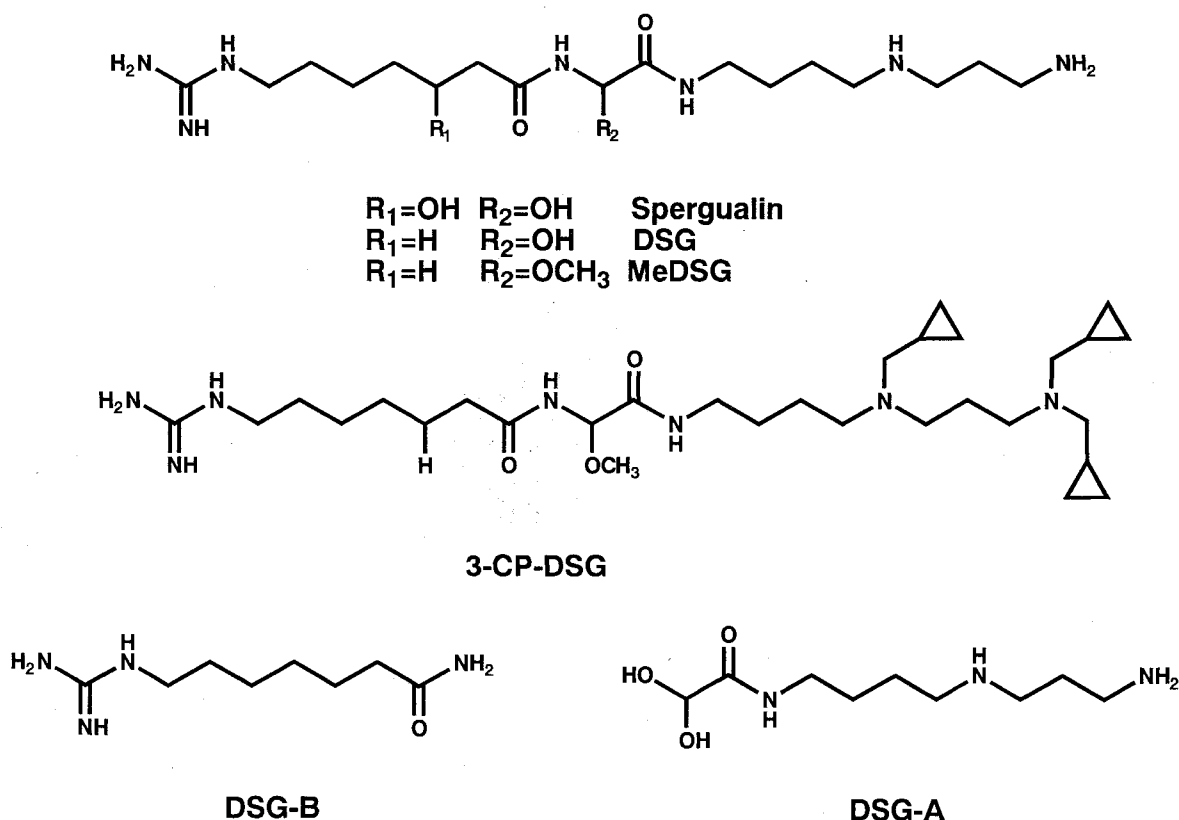
Putrescine, spermidine, spermine, and aminoguanidine

were from Sigma (St. Louis, MO). DL- $\alpha$ -Difluoromethylornithine (DFMO) was from Calbiochem (La Jolla, CA). IR spectra were recorded on a Hitachi 260-10 spectrometer. <sup>1</sup>H NMR spectra were measured with a JEOL JNM A400 spectrometer. HRFAB-MS spectra were measured with a VG AutoSpec mass spectrometer.

#### Preparation of DSG Derivatives

DSG and decomposed elements of DSG (DSG-A and DSG-B) were provided from Takara Shuzou Co. Ltd. (Japan) and Nippon Kayaku Co. Ltd. (Japan), respectively. Methyldeoxyspergualin (MeDSG) was prepared by the method of UMEDA *et al.*,<sup>2</sup> as hydrochloride. To obtain *N*-cyclopropylmethylated DSG (3-CP-DSG), triethylamine (27  $\mu\text{l}$ , 0.193 mmol), cyclopropanecarboxaldehyde (21.2  $\mu\text{l}$ , 0.288 mmol) and sodium cyanoborohydride (19.2 mg, 0.288 mmol) were added to a solution of MeDSG (49.3 mg, 0.096 mmol) in MeOH (2 ml) under ice cooling. After stirring for 4 hours at room temperature, the reaction mixture was diluted with water and chromatographed on a CM-Sephadex C-25 ( $\text{Na}^+$ ) column with a linear gradient elution using water and 0.5 M NaCl solution. Fractions containing 3-CP-DSG were collected and evaporated to

Fig. 1. Structures of DSG and related compounds.



dryness. The residue was extracted with MeOH and the MeOH extract was chromatographed on a Sephadex LH-20 column using MeOH as an eluent. Fractions containing 3-CP-DSG were pooled and evaporated to give 22.0 mg (34%) as a syrup. HRFAB-MS  $m/z$  564.4604 ( $M+H$ )<sup>+</sup> calcd. for  $C_{30}H_{58}N_7O_3$   $m/z$  564.4601; IR (KBr) 3250, 2935, 1660, 1525, 1470, 1375, 1350, 1190, 1090  $cm^{-1}$ ; <sup>1</sup>H NMR ( $CD_3OD$ )  $\delta$ : 0.3~0.5 (6H, m, cyclopropyl- $CH_2$ ), 0.6~0.8 (6H, m, cyclopropyl- $CH_2$ ), 1.0~1.2 (3H, m, cyclopropyl-CH), 1.2~2.0 (12H, m,  $CH_2$ ), 2.0~2.4 (4H, m,  $CH_2$ ), 2.7~3.4 (16H, m,  $CH_2$ ), 3.39 (3H, s,  $OCH_3$ ), 5.30 (1H, s, 11-CH).

#### Cells

Mouse EL-4 lymphoma cells were grown in RPMI1640 medium supplemented with 10% fetal bovine serum (FBS; JRH Biosciences, Lenexa, KS), 100 units/ml of penicillin G, and 100  $\mu g/ml$  of streptomycin at 37°C with 5%  $CO_2$ .

#### Cell Growth

Cells were inoculated into 96-well plates at 3000 cells/well and incubated with or without test drugs for 4

days. The growth was determined by using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) as described.<sup>13</sup> In case of 4h-treatment experiment, the cells were incubated with or without test drugs for 4 hours at 37°C, and then the cells were washed twice with the growth medium and further cultured for 4 days. For competitive assay of DSG, test drugs were added to cells 30 minutes before the addition of DSG.

#### Results and Discussion

15-Deoxyspergualin (DSG) has antiproliferative effect on various cancer cell lines.<sup>4,5</sup> Among them, DSG showed a relatively strong effect on hematopoietic malignant cells. In this study we used mouse EL-4 lymphoma cells. As shown in Fig. 2, DSG inhibited the growth of EL-4 cells with an  $IC_{50}$  0.02  $\mu g/ml$ . In contrast, when the cells were treated with DSG for only 4 hours and then washed out the drug, DSG inhibited the growth apparently with an  $IC_{50}$  0.4  $\mu g/ml$ , even though its effect was weakened (Fig. 2). On the other hand, cycloheximide used as a negative control

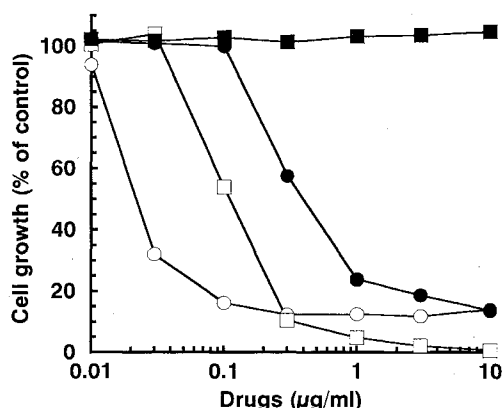
inhibited the growth with an  $IC_{50}$  0.1  $\mu\text{g/ml}$ , but the antiproliferative effect was diminished by 4-hour treatment (Fig. 2). This result shows that the antiproliferative effect of DSG lasts long.

DSG has spermidine and guanidine moieties in its structure (Fig. 1). We next examined the effect of the decomposed elements of DSG. Although one decomposed element containing spermidine moiety (DSG-A) did not affect the growth of EL-4 cells, another element containing guanidine moiety (DSG-B) inhibited it at a high dose of

100  $\mu\text{g/ml}$  (Fig. 3A). Unlike DSG, DSG-B failed to inhibit the growth by only 4-hour treatment (Fig. 3B). Addition of DSG-A along with DSG-B did not enhance the effect of DSG-B (Fig. 3C). We therefore hypothesized that DSG would bind to cells and internalize into the cells through a spermidine moiety and then the internalized DSG would exert its antiproliferative effect through a guanidine moiety.

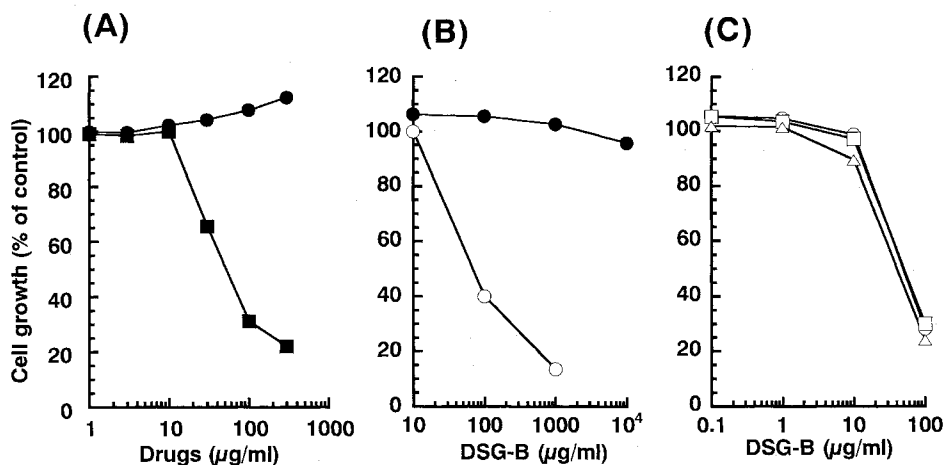
To ascertain the hypothesis, we examined whether the effect of DSG should be abolished by pretreatment of the cells with DSG-A. The cells were preincubated with excess amount of DSG-A for 30 minutes, and then further incubated with DSG along with DSG-A for 4 hours. After washing out the drugs, the cell growth was determined. As a result, pretreatment with DSG-A expectedly diminished the antiproliferative effect of 4-hour treatment DSG dose-dependently (Fig. 4A). As well as DSG-A, pretreatment of the cells with excess amount of polyamines such as putrescine, spermidine, and spermine also abolished the antiproliferative effect of DSG rather strongly (Fig. 4B). The same results were also obtained using spergualin, an original compound of DSG (data not shown). These effects of polyamines were not thought to be due to prevention of polyamine depletion by DSG. It is reported that intracellular polyamines are decreased by DSG treatment.<sup>14)</sup> DL- $\alpha$ -Difluoromethylornithine (DFMO), an inhibitor of ornithine decarboxylase, also decreases the intracellular polyamines.<sup>15)</sup> As shown in Fig. 5, DFMO inhibited the growth with an  $IC_{50}$  50  $\mu\text{g/ml}$ , but the continuous presence of polyamines along with DFMO throughout cultured days diminished the antiproliferative effect of DFMO. This result

Fig. 2. Effect of DSG on EL-4 cell growth.



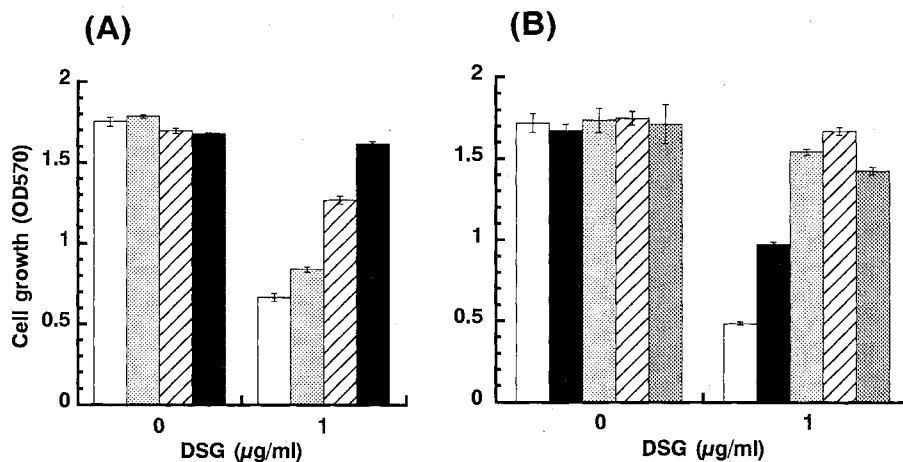
Open symbols, EL-4 cells were cultured with DSG (○) or cycloheximide (□) for 4 days. Closed symbols, the cells were preincubated with DSG (●) or cycloheximide (■) for 4 hours at 37°C, washed and then further cultured for 4 days.

Fig. 3. Effect of DSG decomposed elements on EL-4 cell growth.



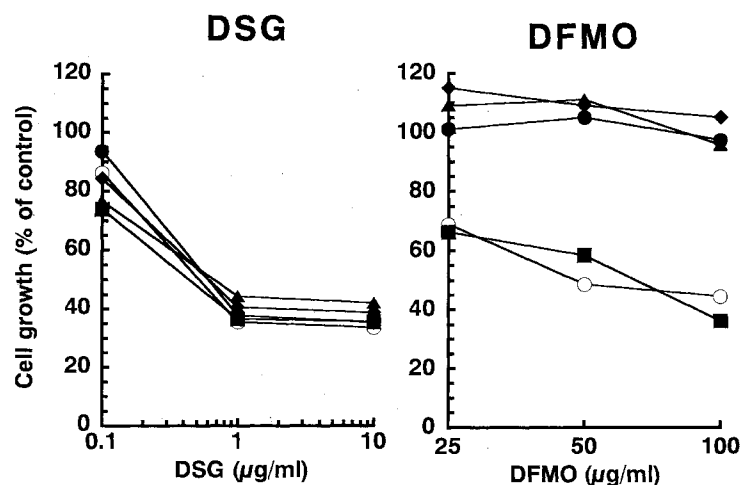
(A) EL-4 cells were cultured with DSG-A (●) or DSG-B (■) for 4 days. (B) The cells were cultured for 4 days with DSG-B (○, continuous; ●, 4-h treatment) as described in Fig. 2. (C) The cells were cultured with the indicated concentrations of DSG-B in the co-presence of DSG-A at 0 (○), 10 (□), and 100 (△)  $\mu\text{g/ml}$  for 4 days.

Fig. 4. Reversal of DSG effect by DSG-A and polyamines.



EL-4 cells were preincubated with DSG-A or polyamines for 30 minutes, and then further incubated with DSG at 1  $\mu\text{g/ml}$  along with DSG-A or polyamines for 4 hours at 37°C. The cells were washed and cultured for 4 days. (A) DSG-A was added at 0 ( $\square$ ), 10 ( $\boxtimes$ ), 100 ( $\boxplus$ ), and 1000 ( $\blacksquare$ )  $\mu\text{g/ml}$ . (B) Polyamines were added as follows, putrescine 32  $\mu\text{g/ml}$  (200  $\mu\text{M}$ ,  $\boxtimes$ ), spermidine 51  $\mu\text{g/ml}$  (200  $\mu\text{M}$ ,  $\boxplus$ ), spermine 70  $\mu\text{g/ml}$  (200  $\mu\text{M}$ ,  $\boxtimes$ ), and DSG-A 58  $\mu\text{g/ml}$  (200  $\mu\text{M}$ ,  $\blacksquare$ ).

Fig. 5. Effect of polyamines on the antiproliferative effect of DSG and DFMO.



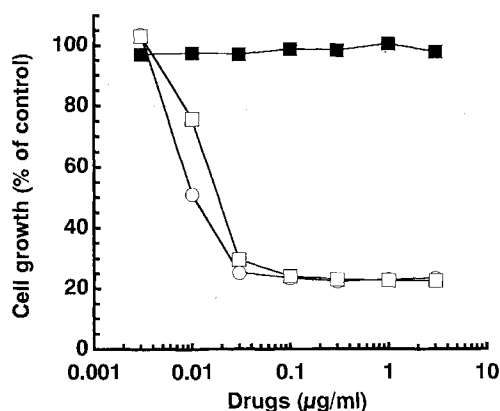
EL-4 cells were preincubated with DSG for 4 hours, washed and then cultured for 4 days in the absence ( $\circ$ ) or the presence of putrescine 100  $\mu\text{g/ml}$  ( $\bullet$ ), aminoguanidine 100  $\mu\text{g/ml}$  ( $\blacksquare$ ), spermidine 10  $\mu\text{g/ml}$  ( $\blacklozenge$ ), or spermine 10  $\mu\text{g/ml}$  ( $\blacktriangle$ ). Because of non-lasting effect of DFMO, DFMO was added along with polyamines without washing. Spermidine and spermine were added with aminoguanidine 100  $\mu\text{g/ml}$  to inhibit oxidation.

indicated that depletion of the intracellular polyamines was prevented by the extra-addition of polyamines. On the other hand, when the cells were preincubated with DSG for 4 hours and then polyamines were added after washing out DSG, the antiproliferative effect of DSG was not affected by the continuous presence of polyamines. Therefore, the preventing effect of polyamines on 4h-treated DSG action

in Fig. 4 was considered to be direct interaction between them. It is reported that DSG inhibits the spermidine transport through cell membrane.<sup>16)</sup> Therefore, our results supported the idea that DSG is transported through the polyamine transporter.

To explore the further possibility that DSG acts through the spermidine moiety, we prepared a DSG derivative, in

Fig. 6. Effect of DSG and its derivatives on EL-4 cell growth.



EL-4 cells were cultured with DSG (○), MeDSG (□), or 3-CP-DSG (■) for 4 days.

which spermidine moiety was structurally hindered, using methyldeoxyspergualin (MeDSG) (Fig. 1). As well as DSG, MeDSG inhibited the growth of EL-4 cells at  $IC_{50}$  0.02  $\mu\text{g/ml}$ . On the contrary, a DSG derivative, cyclopropylmethylated MeDSG (3-CP-DSG) did not affect the growth (Fig. 6). This result shows that the structure in a spermidine moiety in DSG is important for its antiproliferative effect.

These results therefore suggested that DSG binds to the cells strongly through a spermidine moiety and exerts its long-lasting antiproliferative effect. It is speculated that this unique structural characteristic of DSG would confer the potent antitumor effect *in vivo*. Although we are now studying the precise mechanism for DSG action, it might be one of modifications to create an antitumor compound fusing spermidine and a cytotoxic lead compound.

#### Acknowledgement

We thank Ms. K. MIYAJI for preparation of manuscripts. This work was supported by a Grant-in-Aid for Cancer Research from the Ministry of Education, Science, Sports and Culture of Japan.

#### References

- 1) TAKEUCHI, T.; H. IINUMA, S. KUNIMOTO, T. MASUDA, M. ISHIZUKA, M. TAKEUCHI, M. HAMADA, H. NAGANAWA, S. KONDO & H. UMEZAWA: A new antitumor antibiotic, spergualin: Isolation and antitumor activity. *J. Antibiotics* 34: 1619~1621, 1981

- 2) UMEDA, Y.; M. MORIGUCHI, H. KURODA, T. NAKAMURA, H. IINUMA, T. TAKEUCHI & H. UMEZAWA: Synthesis and antitumor activity of spergualin analogues. I. Chemical modification of 7-guanidino-3-hydroxyacyl moiety. *J. Antibiotics* 38: 886~898, 1985
- 3) IWASAWA, H.; S. KONDO, D. IKEDA, T. TAKEUCHI & H. UMEZAWA: Synthesis of (-)-15-deoxyspergualin and (-)-spergualin-15-phosphate. *J. Antibiotics* 35: 1665~1669, 1982
- 4) PLOWMAN, J.; J. STEADMAN, D. HARRISON, M. W. TRADER, J. DANIEL, P. GRISWOLD, M. CHADWICK, M. F. MCCOMISH, D. M. SILVEIRA & D. ZAHARKO: Preclinical antitumor activity and pharmacological properties of deoxyspergualin. *Cancer Res.* 47: 685~689, 1987
- 5) NISHIKAWA, K.; C. SHIBASAKI, M. HIRATSUKA, M. ARAKAWA, K. TAKAHASHI & T. TAKEUCHI: Antitumor spectrum of deoxyspergualin and its lack of cross-resistance to other antitumor agents. *J. Antibiotics* 44: 1101~1109, 1991
- 6) NEMOTO, K.; M. HAYASHI, F. ABE, T. NAKAMURA, M. ISHIZUKA & H. UMEZAWA: Immunosuppressive activities of 15-deoxyspergualin in animals. *J. Antibiotics* 40: 561~562, 1987
- 7) MASUDA, T.; S. MIZUTANI, M. IJIMA, H. ODAI, H. SUDA, M. ISHIZUKA, T. TAKEUCHI & H. UMEZAWA: Immunosuppressive activity of 15-deoxyspergualin and its effect on skin allografts in rats. *J. Antibiotics* 40: 1612~1618, 1987
- 8) NADLER, S. G.; M. A. TEPPER, B. SCHACTER & C. E. MAZZUCCO: Interaction of the immunosuppressant deoxyspergualin with a member of the Hsp70 family of heat shock proteins. *Science* 258: 484~486, 1992
- 9) NISHIKAWA, K.; C. SHIBASAKI, T. UCHIDA, K. TAKAHASHI & T. TAKEUCHI: The nature of *in vivo* cell-killing of deoxyspergualin and its implication in combination with other antitumor agents. *J. Antibiotics* 44: 1237~1246, 1991
- 10) HIRATSUKA, M.; H. KURAMOCHI, K. TAKAHASHI, T. TAKEUCHI & M. OSHIMURA: Cytostatic effect of deoxyspergualin on a murine leukemia cell line L1210. *Jpn. J. Cancer Res.* 82: 1065~1068, 1991
- 11) OIKAWA, T.; M. SHIMAMURA, H. ASHINO-FUSE, T. IWAGUCHI, M. ISHIZUKA & T. TAKEUCHI: Inhibition of angiogenesis by 15-deoxyspergualin. *J. Antibiotics* 44: 1033~1035, 1991
- 12) OIKAWA, T.; M. HASEGAWA, I. MORITA, S.-I. MUROTA, H. ASHINO, M. SHIMAMURA, A. KIUE, R. HAMANAKA, M. KUWANO, M. ISHIZUKA & T. TAKEUCHI: Effect of 15-deoxyspergualin, a microbial angiogenesis inhibitor, on the biological activities of bovine vascular endothelial cells. *Anti-Cancer Drugs* 3: 293~299, 1992
- 13) FUKAZAWA, H.; S. MIZUNO & Y. UEHARA: A microplate assay for quantitation of anchorage-independent growth of transformed cells. *Anal. Biochem.* 228: 83~90, 1995
- 14) HIBASAMI, H.; T. TSUKADA, R. SUZUKI, K. TAKANO, S. TAKAJI, T. TAKEUCHI, S. SHIRAKAWA, T. MURATA & K. NAKASHIMA: 15-Deoxyspergualin, an antiproliferative agent for human and mouse leukemia cells shows inhibitory effects on the synthetic pathway of polyamines. *Anticancer Res.* 11: 325~330, 1991
- 15) WEEKS, C.; A. HERRMANN, F. NELSON & T. SLAGA:  $\alpha$ -Difluoromethylornithine, an irreversible inhibitor of ornithine decarboxylase, inhibits tumor promoter-

induced polyamine accumulation and carcinogenesis in mouse skin. Proc. Natl. Acad. Sci. USA. 79: 6028~6032, 1982

16) KUNIMOTO, S.; C. NOSAKA, C. XU & T. TAKEUCHI: Serum

effect on cellular uptake of spermidine, spergualin, 15-deoxyspergualin, and their metabolites by L5178Y cells. J. Antibiotics 42: 116~122, 1989